

## SHORT COMMUNICATION

UDC 577.112

doi: <https://doi.org/10.15407/ubj90.03.094>**ADAPTOR PROTEIN Ruk/CIN85 MODULATES RESISTANCE TO DOXORUBICIN OF MURINE 4T1 BREAST CANCER CELLS**

I. R. HORAK, D. S. GERASHCHENKO, L. B. DROBOT

*Palladin Institute of Biochemistry, National Academy of Sciences of Ukraine, Kyiv;  
e-mail: iryna.horak@gmail.com*

*The acquisition of chemoresistance in the course of tumor progression includes activation of membrane ABC transporters, detoxification enzymes, cell cycle deceleration and activation of specific signaling pathways such as Akt/mTOR, MAPK, NF- $\kappa$ B. Adaptor proteins play an essential role in the assembly of supra-molecular signaling complexes, maintaining and directing the intracellular signaling. One of such proteins, called Ruk/CIN85, is strongly associated with malignant transformation and metastasis. In present study we investigated the Ruk/CIN85 effect of up/down-regulation on the transforming potential and doxorubicin resistance of highly aggressive mouse breast adenocarcinoma 4T1 cells. It was demonstrated that 4T1 cells overexpressing Ruk/CIN85 possessed increased resistance to doxorubicin (in the range of concentrations 0.1–10.0  $\mu$ M) while knockdown cells were the most sensitive. Also, high levels of Ruk/CIN85 in 4T1 cells positively correlated with their ability to form colonies in semi-solid agar. Ruk/CIN85-overexpressing cells formed four times more colonies in comparison with Ruk/CIN85 knockdown cells, the growth of which revealed higher resistance to doxorubicin action.*

*Key words: Breast cancer, chemoresistance, adaptor proteins, Ruk/CIN85.*

Cancer treatment technics evolved a lot during last decades, and such approaches as personalized medicine [1], immunotherapy [2], cancer stem cells-directed therapy [3], miRNA-directed therapy [4], as well as various ways of targeted drugs delivery [5] have become widespread. However, the development of resistance to the action of antitumor drugs in the course of cancer progression remains the most challenging. Currently, this phenomenon is explained by the heterogeneity of tumor cells including the presence of a pool of cancer stem cells in tumor mass as well as the activation of molecular mechanisms underlying cell plasticity and enabling tumor to develop new options in drugs resistance [6, 7]. Modern studies are focused on targeting the molecular mechanisms, which provide acquired therapeutic resistance of tumor cells: growth factor receptor (RTK) signaling, cell survival regulation, membrane transporters and detoxification enzymes, EMT- and stemness-maintaining signaling [8, 9]. Adaptor/scaf-

fold proteins consisting of domains and motifs involved in intermolecular interactions function as organizers of multimolecular signaling complexes that possess the ability to regulate their composition in time and space thereby directing intracellular signaling. A member of (Ruk/CIN85)/(CD2AP/CMS) family of adaptor proteins, Ruk (regulator of ubiquitous kinase) in rodents and CIN85 (Cbl-interacting protein with MW 85 kDa) in human (thereafter, Ruk/CIN85) [10], consists of three SH3 domains on the N-terminus, proline- and serine-rich domains, and C-terminal coiled-coil region [11, 12]. Due to interaction with numerous binding partners, Ruk/CIN85 is involved in such processes as vesicle-mediated endocytosis and sorting of activated RTKs [13-15], apoptosis [16], cell adhesion, motility and invasiveness [17-19]. Also, the previous studies demonstrated that Ruk/CIN85 is overexpressed in various cancers and may be associated with increased metastatic potential [20-22].

The aim of present study was to investigate the effect of Ruk/CIN85 overexpression and down-regulation on doxorubicin resistance of mouse breast adenocarcinoma 4T1 cells and their transforming potential.

### Materials and Methods

*Cell culture and generation of 4T1 sublines with overexpression and down-regulation of Ruk/CIN85.* Murine breast adenocarcinoma 4T1 cells with different levels of adaptor protein Ruk/CIN85 content were cultured in RPMI-1640 medium (Gibco) supplemented with 10% fetal calf serum (HyClone), 2 mM L-glutamine, 100 U/ml penicillin, 100 µg/ml streptomycin (Gibco), in a humidified atmosphere containing 5% CO<sub>2</sub> at 37 °C. Ruk/CIN85-overexpressing subline RukUp and corresponding control subline Mock were obtained by Ca-phosphate transfection of 4T1 cells with pRc/CMV2-Ruk1 or empty vector, respectively [23]. Ruk/CIN85 expression was suppressed in subline called RukDown using lentiviral construction pLKO.1-shRuk/CIN85 R22 [24] and for generation of control subline (Scr) nontargeting vector was used. Transfected/infected cells were subcloned and selected with 1mg/ml geneticin (G418) or 10 µg/ml puromycin, respectively.

*MTT assay and doxorubicin IC<sub>50</sub> determination.* 4T1 sublines viability after doxorubicin treatment was measured by the MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide) reduction assay. Cells were plated on 96-well plate (10<sup>4</sup> cells per well), allowed them to adhere for 12 h and then treated for 24 hours with 0, 0.1, 1, and 10 µM of doxorubicin. Formation of MTT-formazane was measured spectrophotometrically at wavelength of 570/630 nm using an absorbance microplate reader µQuant (BioTEK). The graph of survival dependence on doxorubicin concentration was built in order to determine doxorubicin IC<sub>50</sub> for each 4T1 subline.

*Soft agar colony formation assay.* 4T1 cells (1x10<sup>3</sup>) were suspended in 150 µl top agar (RPMI supplied with 0.4% agar) with 0.01, 0.05, 0.1 µM of doxorubicin or without it, and layered over 100 µl bottom agar (RPMI supplied with 0.8% agar) in the wells of 96-well plate. After 3 weeks, the cells were photographed with magnification 40x and 200x and the number of colonies was counted. Independent experiments were performed in triplicates.

*Statistical analysis.* All the experiments were independently repeated at least three times, the results were presented as mean ± SEM. Statistical

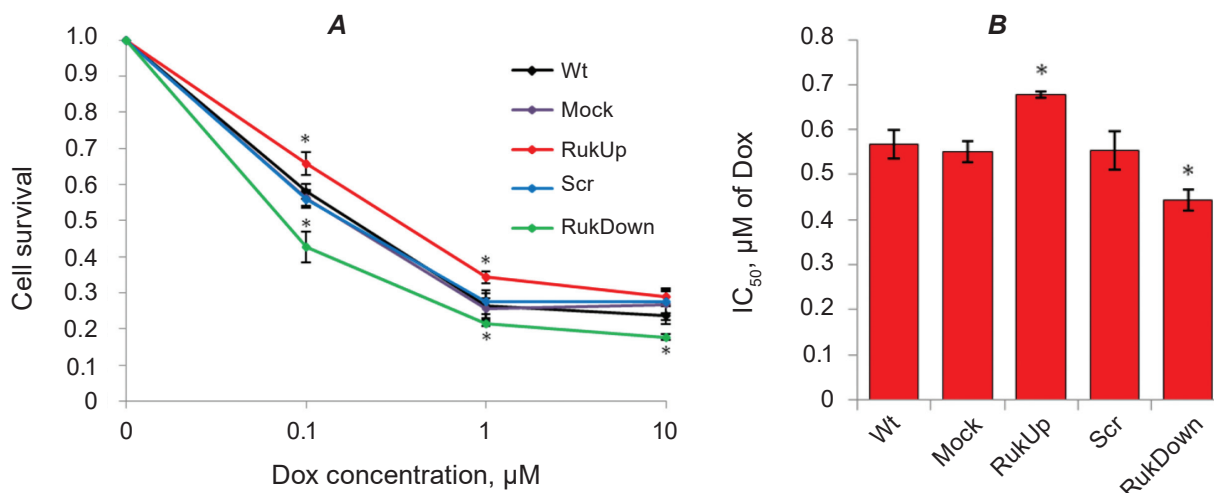
analysis was performed using Statistica software. Data were analysed using factorial ANOVA with Newman-Keuls post-hoc correction for multiple comparisons. Pairwise comparisons were then analyzed by Student's *t*-test for unequal variances and difference between groups was decided to be significant at  $P < 0.05$ .

### Results and Discussion.

In order to study the effect of adaptor protein Ruk/CIN85 on chemoresistance of mouse breast adenocarcinoma 4T1 cells we generated Ruk/CIN85-overexpressing subline RukUp (and corresponding control subline named Mock) using calcium-phosphate transfection as well as Ruk/CIN85-nockdown subline (RukDown) by infection of 4T1 cells with Ruk/CIN85-specific shRNA lentiviral particles (and a control Scr subline, infected with non-targeted virus). As additional control we also used WT 4T1 cells.

Survival of 4T1 sublines with different levels of Ruk/CIN85 expression in the presence of increasing concentrations of doxorubicin (Dox) was evaluated by MTT-test. It was demonstrated, that overexpression of adaptor protein Ruk/CIN85 was associated with significantly increased survival of 4T1 cells at 0.1 µM and 1 µM Dox compared to Mock cells, while down-regulation of Ruk/CIN85- resulted in significantly lower survival rate at 0.1, 1 and 10 µM of Dox in comparison to Scr subline (Fig. 1, A). Then, we evaluated Dox IC<sub>50</sub> for each of the analyzed 4T1 cells sublines and found that the highest value of IC<sub>50</sub> (0.678884 ± 0.006931 µM of Dox) was characteristic of RukUp subline, whereas IC<sub>50</sub> for RukDown cells was the lowest (0.443734 ± 0.02338 µM of Dox) (Fig. 1, B). These results demonstrate that adaptor protein Ruk/CIN85 regulates resistance of 4T1 cells to doxorubicin in concentration-dependent manner: increases resistance in the case of overexpression and attenuates in the case of down-regulation.

Colony formation in soft agar reflects the anchorage-independent growth ability and is widely used as a test for malignant transformation of tumor cells [25]. We analyzed the colony formation potential of 4T1 cells with up- or down-regulation of Ruk/CIN85 in the presence of 0.01, 0.05, and 0.1 µM doxorubicin. It was demonstrated that under the control conditions (without Dox) RukUp cells formed significantly higher number of colonies in semi-solid agar in comparison to Mock cells. In contrary, down-regulation of Ruk/CIN85 led to suppression of colony



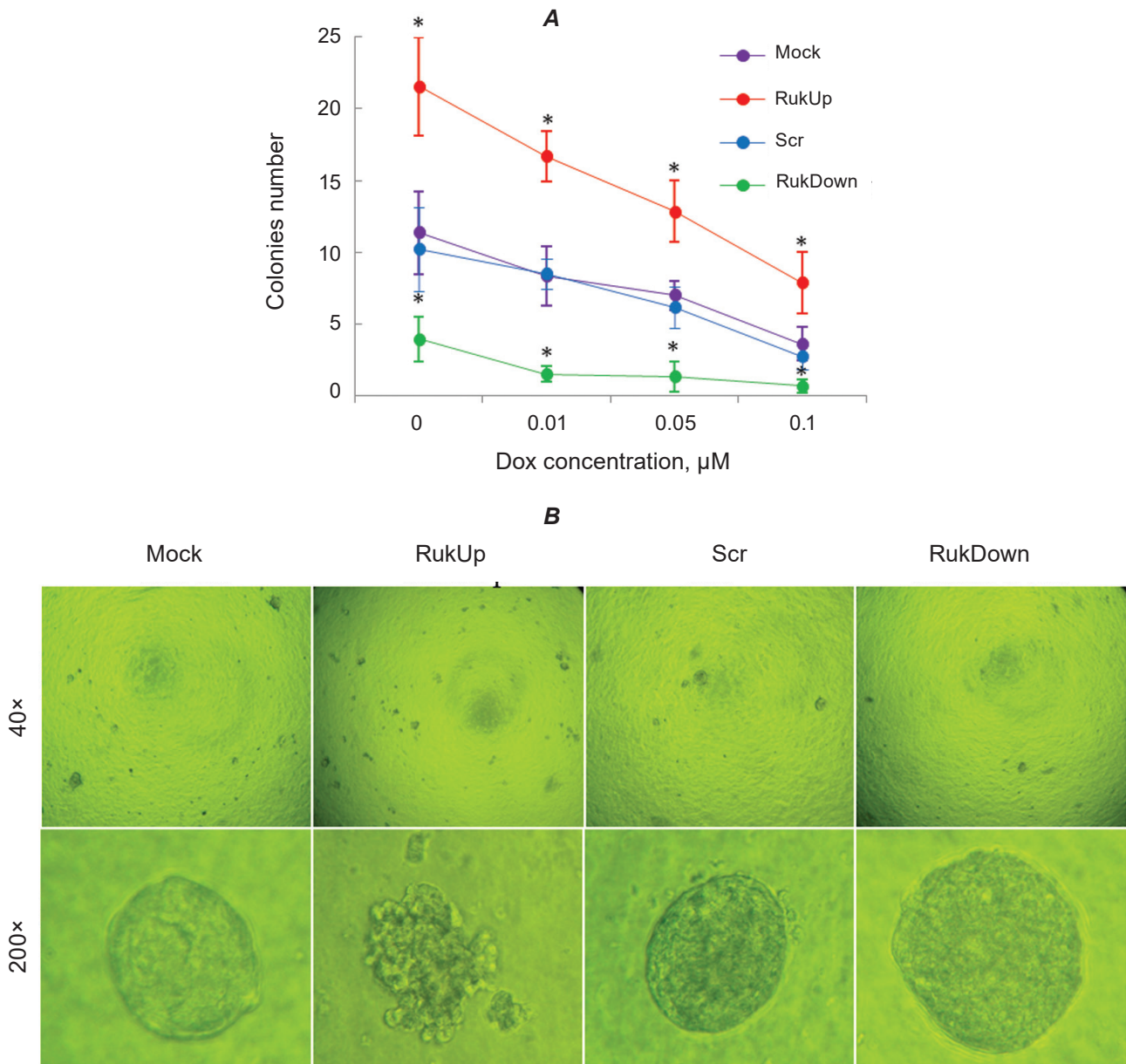
**Fig. 1.** High *Ruk/CIN85* expression level in 4T1 cells positively correlates with doxorubicin resistance. **A** – Doxorubicin survival of mouse breast adenocarcinoma 4T1 cells with different levels of *Ruk/CIN85* expression (*Wt* – wild-type 4T1 cells; *Mock* – 4T1 cells transfected with empty *pRc/CMV2* plasmid; *RukUp* – 4T1 cells, stably transfected with *pRc/CMV2* plasmid encoding full-length form of *Ruk/CIN85*; *Scr* – 4T1 cells infected with irrelevant lentivirus; *RukDown* – 4T1 cells stably infected with *Ruk/CIN85*-specific shRNA lentivirus). **B** – *Ruk/CIN85* effect on doxorubicin  $IC_{50}$  value of 4T1 cells. \*  $P < 0.05$  in comparison to corresponding control

formation ability. In the presence of Dox the overall number of colonies formed by each subline was decreased in comparison to control without Dox. Importantly, at each Dox concentration point we found significantly higher number of colonies for *RukUp* subline in comparison to *Mock* cells, and substantial decrease of colonies number formed by *RukDown* cells in comparison to *Scr* control (Fig. 2, *A*). The representative images of colonies characteristic of each 4T1 subline are presented at Fig. 2, *B*. It should be noticed that *RukDown* cells formed big, spherical colonies with smooth edges, whereas colonies formed by control cells had also spherical shape but were something smaller in size. In contrast, *Ruk/CIN85*-overexpressing cells gave colonies of smaller size with irregular edges and wherein small colonies usually surrounded bigger one. Such features in the morphology of soft agar colonies suggest that up-regulation of *Ruk/CIN85* in 4T1 cells drive the development of a more malignant cellular phenotype potentially associated with increased metastatic potential.

Chemoresistance is an essential feature of cancer stem cells (CSCs) – small subpopulation of tumor cells with self-renewal and tumorigenic potential that allow survival and spreading of the tumor [26]. We demonstrated that expression level of *Ruk/CIN85* correlates with the ability of 4T1 cells to form colonies in soft agar and the ability of these

colonies to survive in the presence of doxorubicine. Taking into account that *Ruk/CIN85* overexpression lead to the increased malignant properties also on other cell types [28, 28], and its ability to affect cell cycle progression and proliferation [17, 22], it may be concluded, that *Ruk/CIN85*-overexpressing cells are enriched with subpopulation with the properties of CSCs.

Currently, it is known that different mechanisms could be responsible for the development of drug resistance including those involved in the control of cell cycle progression and DNA damage response, providing evasion of apoptosis and increased drug efflux (such as ATP-binding cassette (ABC) membrane transporters) or detoxification of drugs (aldehyde dehydrogenase ALDH) [29]. In addition, acquisition of drug resistant phenotype requires activation of PI3K/Akt/mTOR, NF- $\kappa$ B, p53 pathways [30-32]. Previous studies demonstrated, that overexpression of adaptor protein *Ruk/CIN85* in another breast cancer model (human breast adenocarcinoma MCF-7 cells) was accompanied by increased resistance to cisplatin and etoposide along with increased rhodamine 123 efflux and ALDH activity [32], that is consistent with our results on 4T1 breast cancer cells. Moreover, *Ruk/CIN85*-overexpressing MCF-7 cells were also demonstrated to have constitutively activated Akt kinase [22] that plays major role in the chemoresistance mechanisms.



**Fig. 2.** Adaptor protein Ruk/CIN85 modulates the transforming potential of 4T1 cells in a manner dependent on its expression levels. **A** – Soft agar colonies formation ability of 4T1 cells with different levels of Ruk/CIN85 expression levels in the presence or absence of doxorubicin (Wt – wild-type 4T1 cells; Mock – 4T1 cells transfected with empty pRc/CMV2 plasmid; RukUp – 4T1 cells, stably transfected with pRc/CMV2 plasmid encoding full-length form of Ruk/CIN85; Scr – 4T1 cells infected with irrelevant lentivirus; RukDown – 4T1 cells stably infected with Ruk/CIN85-specific shRNA lentivirus). \*  $P < 0.05$  in comparison to corresponding control. **B** – Representative pictures of soft agar colonies formed by 4T1 cells with different levels of Ruk/CIN85 expression

In the same time, doxorubicin-resistant breast cancer MCF-7 cells have considerably repressed TOP2A gene, the product of which is the main target of doxorubicin, as well as up-regulated expression of ABC membrane transporters, cell cycle and proliferation regulators [33]. Along with the known information

regarding the complexity and diversity of chemoresistance mechanisms, our data suggest that adaptor protein Ruk/CIN85 may function as important component of regulatory networks need to acquire drug resistant phenotype by breast cancer cells. This means that Ruk/CIN85-overexpressing breast cancer



cells acquire drug resistant phenotype, provided by various molecular mechanisms.

In this study we analysed the effect of overexpression/down-regulation of adaptor protein Ruk/CIN85 on the survival and soft agar colonies formation ability of mouse breast adenocarcinoma 4T1 cells in the presence of doxorubicin. We found that increased expression level of Ruk/CIN85 positively correlates with doxorubicin resistance and transforming potential of 4T1 cells.

This study was supported by SCOPES grant No IZ73ZO supported by Swiss National Science Foundation SNSF.

### **АДАПТЕРНИЙ ПРОТЕЇН Ruk/CIN85 МОДУЮЄ РЕЗИСТЕНТНІСТЬ КЛІТИН АДЕНОКАРЦИНОМИ МОЛОЧНОЇ ЗАЛОЗИ МИШІ ЛІНІЇ 4T1 ДО ДОКСОРУБІЦИНУ**

*I. P. Horak, D. S. Geraщенко, L. B. Drobot*

Інститут біохімії ім. О. В. Палладіна  
НАН України, Київ;  
e-mail: iryna.horak@gmail.com

Набуття пухлинними клітинами хіміорезистентності відбувається завдяки активації мембранних АВС-транспортів, ензимів деградації ксенобіотиків, затримці клітинного циклу та активації специфічних сигнальних шляхів, таких як Akt/mTOR, MAPK, NF-κB. Адаптерні протеїни відіграють важливу роль у збиранні надмолекулярних сигнальних комплексів, підтриманні та спрямуванні внутрішньоклітинного сигналювання. Один з таких протеїнів, Ruk/CIN85, залучений до процесів злоякісної трансформації та метастазування. В цій роботі досліджено вплив адаптерного протеїну Ruk/CIN85 на трансформувальний потенціал та резистентність до доксорубіцину клітин аденокарциноми молочної залози миші лінії 4T1. Продемонстровано, що Ruk/CIN85 модулює резистентність клітин 4T1 до доксорубіцину в концентрації 0,1–10,0 μM. Також виявлено позитивний зв'язок між вмістом Ruk/CIN85 у клітинах лінії 4T1 та здатністю до формування колоній у напіврідкому агарі, в тому числі в присутності 0,01–0,1 μM доксорубіцину.

**Ключові слова:** рак молочної залози, хіміорезистентність, адаптерні протеїни, Ruk/CIN85.

### **АДАПТЕРНЫЙ ПРОТЕИН Ruk/CIN85 МОДУЛИРУЕТ РЕЗИСТЕНТНОСТЬ КЛЕТОК АДЕНОКАРЦИНОМЫ МОЛОЧНОЙ ЖЕЛЕЗЫ МЫШИ ЛИНИИ 4T1 К ДОКСОРУБИЦИНУ**

*I. P. Horak, D. S. Geraщенко, L. B. Drobot*

Інститут біохімії ім. А. В. Палладіна  
НАН України, Київ;  
e-mail: iryna.horak@gmail.com

Приобретение опухолевыми клетками химиорезистентности происходит за счет активации мембранных АВС-транспортёров, энзимов деградации ксенобіотиков, задержки клеточного цикла и активации специфических сигнальных путей, таких как Akt/mTOR, MAPK, NF-κB. Адаптерные протеины играют важную роль в сборке надмолекулярных сигнальных комплексов, поддержке и направленности внутриклеточной сигнализации. Один из таких протеинов, Ruk/CIN85, вовлечен в процессы злокачественной трансформации и метастазирования. В данной работе исследовано влияние адаптерного протеина Ruk/CIN85 на трансформирующий потенциал и резистентность к доксорубіцину клеток аденокарциномы молочной железы мыши линии 4T1. Продемонстрировано, что Ruk/CIN85 модулирует резистентность клеток 4T1 к доксорубіцину в концентрации 0,1–10,0 μM. Также выявлена положительная связь между содержанием Ruk/CIN85 в клетках линии 4T1 и способностью к формированию колоний в полужидком агаре, в том числе в присутствии 0,01–0,1 μM доксорубіцина.

**Ключевые слова:** рак молочной железы, химиорезистентность, адаптерные протеины, Ruk/CIN85.

#### **References**

1. Jackson SE, Chester JD. Personalised cancer medicine. *Int J Cancer*. 2015; 137(2): 262-266.
2. Vanneman M, Dranoff G. Combining immunotherapy and targeted therapies in cancer treatment. *Nat Rev Cancer*. 2012; 12(4): 237-251.
3. Wang T, Shigdar S, Gantier MP, Hou Y, Wang L, Li Y, Shamaileh HA, Yin W, Zhou SF, Zhao X, Duan W. Cancer stem cell targeted therapy: progress amid controversies. *Oncotarget*. 2015; 6(42): 44191-44206.

4. Monroig Pdel C, Chen L, Zhang S, Calin GA. Small molecule compounds targeting miRNAs for cancer therapy. *Adv Drug Deliv Rev.* 2015; 81: 104-116.
5. Mitra AK, Agrahari V, Mandal A, Cholkar K, Natarajan C, Shah S, Joseph M, Trinh HM, Vaishya R, Yang X, Hao Y, Khurana V, Pal D. Novel delivery approaches for cancer therapeutics. *J Control Release.* 2015; 219: 248-268.
6. Fisher R, Pusztai L, Swanton C. Cancer heterogeneity: implications for targeted therapeutics. *Br J Cancer.* 2013; 108(3): 479-485.
7. Garraway LA, Jänne PA. Circumventing cancer drug resistance in the era of personalized medicine. *Cancer Discov.* 2012; 2(3): 214-226.
8. Westover D, Li F. New trends for overcoming ABCG2/BCRP-mediated resistance to cancer therapies. *J Exp Clin Cancer Res.* 2015; 34: 159.
9. Du B, Shim JS. Targeting Epithelial-Mesenchymal Transition (EMT) to Overcome Drug Resistance in Cancer. *Molecules.* 2016; 21(7). pii: E965.
10. Dikic I. CIN85/CMS family of adaptor molecules. *FEBS Lett.* 2002; 529(1): 110-115.
11. Buchman VL, Luke C, Borthwick EB, Gout I, Ninkina N. Organization of the mouse Ruk locus and expression of isoforms in mouse tissues. *Gene.* 2002; 295(1): 13-17.
12. Finniss S, Movsisyan A, Billecke C, Schmidt M, Randazzo L, Chen B, Bögler O. Studying protein isoforms of the adaptor SETA/CIN85/Ruk with monoclonal antibodies. *Biochem Biophys Res Commun.* 2004; 325(1): 174-182.
13. Havrylov S, Ichioka F, Powell K, Borthwick EB, Baranska J, Maki M, Buchman VL. Adaptor protein Ruk/CIN85 is associated with a subset of COPI-coated membranes of the Golgi complex. *Traffic.* 2008; 9(5): 798-812.
14. Schroeder B, Srivatsan S, Shaw A, Billadeau D, McNiven MA. CIN85 phosphorylation is essential for EGFR ubiquitination and sorting into multivesicular bodies. *Mol Biol Cell.* 2012; 23(18): 3602-3611.
15. Ahmad G, Mohapatra BC, Schulte NA, Nadeau SA, Luan H, Zutshi N, Tom E, Ortega-Cava C, Tu C, Sanada M, Ogawa S, Toews ML, Band V, Band H. Cbl-family ubiquitin ligases and their recruitment of CIN85 are largely dispensable for epidermal growth factor receptor endocytosis. *Int J Biochem Cell Biol.* 2014; 57: 123-134.
16. Narita T, Nishimura T, Yoshizaki K, Taniyama T. CIN85 associates with TNF receptor 1 via Src and modulates TNF-alpha-induced apoptosis. *Exp Cell Res.* 2005; 304(1): 256-264.
17. Havrylov S, Redowicz MJ, Buchman VL. Emerging roles of Ruk/CIN85 in vesicle-mediated transport, adhesion, migration and malignancy. *Traffic.* 2010; 11(6): 721-731.
18. Bai SW, Herrera-Abreu MT, Rohn JL, Racine V, Tajadura V, Suryavanshi N, Bechtel S, Wiemann S, Baum B, Ridley AJ. Identification and characterization of a set of conserved and new regulators of cytoskeletal organization, cell morphology and migration. *BMC Biol.* 2011; 9: 54.
19. Yasin HWR, van Rensburg SH, Feiler CE, Johnson RI. The adaptor protein Cindr regulates JNK activity to maintain epithelial sheet integrity. *Dev Biol.* 2016; 410(2): 135-149.
20. Cascio S, Finn OJ. Complex of MUC1, CIN85 and Cbl in colon cancer progression and metastasis. *Cancers (Basel).* 2015; 7(1): 342-352.
21. Ma Y, Ye F, Xie X, Zhou C, Lu W. Significance of PTPRZ1 and CIN85 expression in cervical carcinoma. *Arch Gynecol Obstet.* 2011; 284(3): 699-704.
22. Samoylenko A, Vynnytska-Myronovska B, Byts N, Kozlova N, Basaraba O, Pasichnyk G, Palyvoda K, Bobak Y, Barska M, Mayevska O, Rzhepetsky Y, Shuvayeva H, Lyzogubov V, Usenko V, Savran V, Volodko N, Buchman V, Kietzmann T, Drobot L. Increased levels of the HER1 adaptor protein Ruk1/CIN85 contribute to breast cancer malignancy. *Carcinogenesis.* 2012; 33(10): 1976-1984.
23. Gout I, Middleton G, Adu J, Ninkina NN, Drobot LB, Filonenko V, Matsuka G, Davies AM, Waterfield M, Buchman VL. Negative regulation of PI 3-kinase by Ruk, a novel adaptor protein. *EMBO J.* 2000; 19(15): 4015-4025.
24. Samoylenko AA, Byts NV, Pasichnyk GV, Kozlova NV, Bazalii AV, Gerashchenko DS, Shandrenko SG, Vorotnikov AV, Kietzmann T, Komisarenko SV, Drobot LB. Recombinant lentivirus-mediated silencing of adaptor protein Ruk/CIN85 expression influences biological responses of tumor cells. *Biotechnol Acta.* 2013; 6(4): 182-189.
25. Borowicz S, Van Scoyk M, Avasarala S, Karuppusamy Rathinam MK, Tauler J, Bikkavilli RK, Winn RA. The soft agar colony formation assay. *J Vis Exp.* 2014; (92): e51998.

26. Cojoc M, Mäbert K, Muders MH, Dubrovskaya A. A role for cancer stem cells in therapy resistance: cellular and molecular mechanisms. *Semin Cancer Biol.* 2015; 31: 16-27.
27. Böglér O, Furnari FB, Kindler-Roehrborn A, Sykes VW, Yung R, Huang HJ, Cavenee WK. SETA: a novel SH3 domain-containing adapter molecule associated with malignancy in astrocytes. *Neuro Oncol.* 2000; 2(1): 6-15.
28. Cascio S, Farkas AM, Hughey RP, Finn OJ. Altered glycosylation of MUC1 influences its association with CIN85: the role of this novel complex in cancer cell invasion and migration. *Oncotarget.* 2013; 4(10): 1686-1697.
29. Sokolosky ML, Stadelman KM, Chappell WH, Abrams SL, Martelli AM, Stivala F, Libra M, Nicoletti F, Drobot LB, Franklin RA, Steelman LS, McCubrey JA. Involvement of Akt-1 and mTOR in sensitivity of breast cancer to targeted therapy. *Oncotarget.* 2011; 2(7): 538-550.
30. Niero EL, Rocha-Sales B, Lauand C, Cortez BA, de Souza MM, Rezende-Teixeira P, Urabayashi MS, Martens AA, Neves JH, Machado-Santelli GM. The multiple facets of drug resistance: one history, different approaches. *J Exp Clin Cancer Res.* 2014; 33: 37.
31. McCubrey JA, Abrams SL, Fitzgerald TL, Cocco L, Martelli AM, Montalto G, Cervello M, Scalisi A, Candido S, Libra M, Steelman LS. Roles of signaling pathways in drug resistance, cancer initiating cells and cancer progression and metastasis. *Adv Biol Regul.* 2015; 57: 75-101.
32. Pasichnyk GV, Povorozniuk OO, Horak IR, Gerashchenko DS, Ponomarenko OV, Samoylenko AA, Byts NV, Drobot LB. Overexpression of adaptor protein Rukl/CIN85 in human breast adenocarcinoma cell line MCF-7 is accompanied by increased chemoresistance. *Rep Nat Acad Sci Ukraine.* 2013; 12: 149-156. (In Ukrainian).
33. AbuHammad S, Zihlif M. Gene expression alterations in doxorubicin resistant MCF7 breast cancer cell line. *Genomics.* 2013; 101(4): 213-220.

Received 31.12.2017